Abstract. Forkhead Box M1 (FoxM1) is one of the most important oncogenes, and overexpression of FoxM1 has been reported in many cancers, including colon cancer. In the present study, the authors attempted to reveal the mechanism underlying its effects on proliferation through autophagy in the sw480 cell line. FoxM1 is knocked down through short hairpin (sh)RNA in the sw480 cell line. A series of experiments were conducted to examine its function on proliferation and LC3 and P62 were used to measure level of autophagy. Autophagy in the shFoxM1 cell was demonstrated as significantly inhibited compared with the negative control. Additional auto-flux was also tested, downregulation of FoxM1 served the same role as BA1 in autophagy. Furthermore, downregulating FoxM1 inhibited cell proliferation in the sw480 cell line.

Introduction

Colon cancer is the third most frequently diagnosed cancer and one of the leading causes of cancer-related deaths worldwide (1,2). Many studies have attempted to demonstrate its effects on tumorigenesis, however, further investigation is still required.

Forkhead Box M1 (FoxM1), a member of the Fox protein family characterized by a conserved winged-helix DNA binding domain, promotes cell cycle progression by inducing both transition from G1 to S phase, and transition from G2 to M phase (3). Previous studies demonstrated that overexpression of FoxM1 is observed in a wide variety of cancers, including breast, ovarian, colon, liver, pancreatic, cervical and gastric cancers (4-6). Moreover, expression of FoxM1 is correlated with a clinically aggressive, drug-resistant, cancer phenotype and poor patient survival in many cancers (7). FoxM1 is regulated by oncogenic signals, including many crucial factors, such as the p53 tumor suppressor protein, which is mutated in half of human cancers (8). FoxM1 is considered to be one of the most important targets within the oncogenic pathway. Many studies of FOXM1 in the tumorigenesis of colon cancer have been conducted, more detailed research is needed, as its relation to autophagy is still unclear.

Autophagy was first reported by Christian de Duve at the International Lysosomal Conference in 1963. It refers to how some protein and organelles were packed into lysosome to degrade, and that the degradation products may be recycled to produce energy. It is widely accepted that autophagy is a means of self-protection when the cell is near death (9,10). The autophagy process is a series of gradual evolution of autophagy structure. Once the autophagy is induced, the isolation membrane and phagophore are formed to combine with the protein and organelle. The isolation membrane extends and wraps the enclosed cytoplasmic component to form a bilayer membrane. Structure-autophosome, autophosome combines with lysosome to form autopholysome where the enclosed cytoplasmid is degraded (9,10).

Materials and methods

Reverse transcription-quantitative polymerase chain reaction. A total of five frozen tumors and paired normal tissues stored at -80°C are enrolled to test the expression level of
FoxM1 mRNA, β-actin was used as a control. Total RNA was extracted with TRIzol reagent, reverse transcription was performed using oligo dT primers and SuperScript III RT (Invitrogen; Thermo Fisher Scientific, Inc., Waltham, MA, USA).

**Cell lines, culture conditions.** The human sw480 cell line was obtained from the American Type Culture Collection (Manassas, VA, USA). All cell lines were cultured in RPMI-1640 Medium supplemented with 10% fetal bovine serum. Cells were cultured at 37°C in a humidified incubator with 5% CO₂.

**Lentivirus production.** Short hairpin (sh)RNA and a negative control were transfected into HEK293T by Lipofectamine® 3000 to produce the lentivirus following manufacturer's protocol. The shRNA and negative control were transfected into HEK293T cells when the cell density was 95-99%. The medium was collected following incubating for 72 h at 37°C in a humidified incubator with 5% CO₂. sw480 cells were seeded into a six-well plate, and, following incubating for 12 h, 1 ml lentivirus medium with 1 ml complete medium was added into the well. Following incubating for 24 h, the media were replaced with complete medium. After another 72 h incubation, a stable cell line was established.

**Infection with shRNA.** FoxM1 shRNA was purchased from Sigma-Aldrich; Merck KGaA. Darmstadt, Germany (sequence, 5'-GGAAATGCTTGTGATTCAACA-3'). The lentivirus was used to transfect shFoxM1 into the sw480 cell line. Exponentially growing untreated cells were plated 24 h prior to infection. Plated cells were infected with FoxM1 shRNA or a negative control with 1 ml of the lentivirus for 72 h. Fluorescence microscopy and western blotting were used to ensure that the shRNA was successfully transfected into the sw480 cell line, and that FoxM1 was successfully knocked down.

**Western blot analysis.** Cells were seeded in 25 cm² culture flasks (2.5x10⁵ cells/5 ml medium). Following treatment, the cells were collected, washed twice in ice-cold phosphate-buffered saline and lysed in a lysis buffer at 4°C. The total protein concentration for each sample was determined with a bicinchoninic acid kit. Western blotting was performed with 40 µg for each sample by SDS-PAGE and transferred onto polyvinylidene fluoride transfer membranes for western blotting. The membranes were blocked with a blocking buffer (5% dry milk in TBS-Tween-20) for 1 h. After being washed with TBS-T, the membranes were probed with the following primary antibodies: FoxM1, Lc3, P62, p-H2A.X and β-actin. Following another wash with TBS-T, the membranes were incubated with anti-rabbit or anti-mouse secondary antibody. All antibodies were diluted in TBS-T containing 5% BSA. Chemiluminescence detection was performed with X-ray film.

**Cell viability and proliferation assays.** Cell viability and proliferation were measured by MTT. The cells were digested with pancreatin. Following centrifugation, the cells were resuspended into fresh complete culture solution and then seeded into a 96-well plate (5x10³/200 µl), repeating 5-wells. Then, the cells were incubated in an incubator with 5% CO₂ at 37°C for 24-96 h, and 20 µl MTT solution (5 mg/ml) was added for color generation. Incubation was continued for extra 4 h in an incubator with 5% CO₂ at 37°C and the culture solution was aspirated following 4 h. DMSO (200 µl) was added into every well and the samples were slightly shaken for 10 min to speed up the dissolution of crystals. The absorbance value of each well was detected by MTT following 0, 24, 48, 72 and 96 h. The absorbance value and interval time were posited as vertical coordinates and abscissa, respectively,
following which, the MTT graph was drawn. All procedures were repeated three times.

Cell cycle by flow cytometry. The cycle distribution was analyzed by flow cytometry following staining with propidium iodide (PI) solution. Briefly, sw480-shFoxM1 and negative control cells were seeded into a 6-well plate with 3x10^6/well following incubating for 24 h, cells were fixed with 75% ethanol. Next, the cells were incubated with 500 μl solution containing 50 mg/ml PI and 0.1% Triton X-100 in the dark and analyzed by flow cytometry. Finally, the data was analyzed by FlowJob software.

Colony formation assay. Cells were trypsinized and plated in 10 cm dishes at a density of 5x10^4/dish. Following incubating at 37°C 5% CO_2 for 2 weeks, the authors used Methylrosanilinium Chloride Solution to dye colony.

Soft-agar clonogenicity assay. Clonogenic growth of cells was evaluated by seeding 0.5x10^3 cells in 0.5 ml RPMI-1640 supplemented with 0.33% agar and 10% FBS. Cells were grown in vitro (37°C, 5% CO_2) for 3 weeks.

Results

FoxM1 is significantly upregulated in colon carcinoma compared with paired normal tissues. FoxM1 was upregulated in colon cancers, when compared with normal tissues in many published researches at both mRNA and protein level and attributes to the tumorigenesis of colon cancer (Fig. 1).
shFoxM1 and the negative control lentivirus was successfully produced in HEK293T cells and was successfully infected into the sw480 cell line. To investigate the potential function of FoxM1 in the sw480 cell line, the expression level of FoxM1 was knocked down by using a lentivirus system. Following infection for 3 days, after screening with puromycin for 3 days. Fluorescence microscopy was used to detect GFP and the infection efficiency. ~99% cells were successfully infected with lentivirus (Fig. 2).

FoxM1 was successfully knocked down in the sw480 cell line and downregulating FoxM1 significantly reduced autophagy level. Following establishing the stable shFoxM1 cell line, the authors investigated the relationship between downregulating FoxM1 and autophagy level. LC3 p62 and p-H2A.X were tested (Fig. 3A). p62 is a selective substrate for autophagy-lysosome degradation, so, total p62 protein levels reflect autophagic activity. To test whether the decrease in autophagosomes is due to decreased autophagy activity or a block in downstream degradation, the authors performed an autophagic flux assay. Bafilomycin A1 (Baf A1) is a lysosomotropic reagent that blocks autophagosome degradation. As was expected, Baf A1 treatment caused elevated levels of LC3 in the NC, shFoxM1 cells. In addition, p62 protein levels increased upon Baf A1 treatment in the NC, shFoxM1 cells (Fig. 3B). Therefore, it may be concluded that downregulating FoxM1 decreases autophagic activity.

Knockdown of FoxM1 inhibits cell proliferation in the sw480 cell line. Following successfully FoxM1, the authors conducted a series of experiments to test FoxM1-mediated biological function. A clonogenic assay was used to measure the ability to form the foci. Downregulation of FoxM1 resulted in a significantly reduction in the number of colonies compared with the control group (Fig. 4C and D). Flow cytometry was used to test the cell cycle. Downregulation of FoxM1 indicated G1 arrest compared with the negative control (Fig. 4B). MTT were used to examine the effort on the short-term proliferation of downregulated FoxM1. Consistently, downregulated FoxM1 reduced the proliferation in the sw480 cell line in the early stage, which is corresponded with the flow cytometry results (Fig. 4A).

Discussion

Colon cancer is one of the most frequently diagnosed cancers and has been well studied in the past decades. FoxM1, a member of the Fox protein family, characterized by a conserved winged-helix DNA binding domain, promotes cell cycle progression by inducing both transition from G1 to S phase and transition from G2 to M phase. Autophagy is one of the most prominently studied fields worldwide, however, the mechanism remains unclear in the sw480 cell line; the authors aimed to uncover its mechanism in the present study.

A previous study of the authors demonstrated the correlation between FoxM1 expression and autophagy level though detecting LC3B and P62. The ratio of LC3BII to LC3BI increased and P62 significantly accumulated in sw480-shFoxM1, indicating that the autophagy level was declined. Furthermore, Baf A1 is a lysosomotropic reagent that blocks autophagosome degradation. As expected, Baf A1 treatment caused elevated levels of LC3 in the NC, shFoxM1 cells. In addition, p62 protein levels increased upon Baf A1 treatment in the NC, shFoxM1 cells. Therefore, downregulating FoxM1 decreases autophagic activity.

The damage of DNA was enhanced, the accumulated p-H2A.X also confirmed this conclusion. In the current research, it was hypothesized that knockdown of FoxM1 may exert its function by upregulating DNA damage and down-regulating autophagy level through inhibiting the recycling of many organelles to affect tumor tumorigenesis.

In conclusion, the present results demonstrated the relationship between downregulating FoxM1 and autophagy and its potential mechanism. Downregulating FoxM1 significantly reduced autophagy level and enhanced DNA damage to inhibit proliferation in the sw480 cell line.

References